# Phenotypic diversity of wild tomato (Solanum lycopersicum L.) populations 

Alvarado-Rodríguez, Rommel I. ${ }^{1}$; Legaria-Solano, Juan Porfirio ${ }^{2 *}$<br>${ }^{1}$ Postgrado en Biotecnología Agrícola, Universidad Autónoma Chapingo, Carretera Federal México-Texcoco km 38.5, Chapingo, Estado de México, México, C. P. 56230.<br>${ }^{2}$ Universidad Autónoma Chapingo, Carretera Federal México-Texcoco km 38.5, Chapingo, Estado de México, México, C. P. 56230.<br>* Correspondence: legarias.juan@yahoo.com

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#### Abstract

Objective: To produce information about the morpho-agronomic variability of 15 wild tomato populations from different areas of Mexico. Design/Methodology/Approach: A completely experimental design was used, comprised of 17 treatments ( 15 wild tomato populations and two commercial tomato populations) and 10 repetitions (individuals). The experimental unit was a plant (individual) which was subjected to an evaluation of 65 morpho-agronomic descriptors, proposed by Biodiversity International. An analysis of variance using repeated measurements was carried out and the mean differences were compared with Tukey's multiple comparison test ( $\mathrm{p} \leq 0.05$ ). The quantitative and qualitative variables were subjected to a main component and multiple correspondence analyses, respectively. Results: A wide variability of the morphological traits and the quality attributes of the fruits -such as consistency and total soluble solids- was recorded. The main component and multiple correspondence analyses accounted for 67.41 and $42.06 \%$ of the phenotypic variation, respectively, in the first three components and dimensions. The more discriminatory characteristics belonged to fruits and cymes, based on which the populations were divided into four groups. The first group was made up of heirloom tomatoes with multiparous cyme, and red, small, and medium fruits; the second group was made up of cherry and grape tomatoes with uniparous and multiparous cymes, and yellow, orange, red, and very small fruits; the third group was made up of beef and cocktail tomatoes with uniparous and bifurcated cymes and red, orange, yellow, and small and medium fruits; finally, the fourth group was made up of purple beef tomatoes with uniparous cymes and medium size tomatoes. Study Limitations/Implications: A molecular characterization must be carried out in order to better understand the variability of these populations. Findings/Conclusions: All wild tomato populations show a wide genetic heritage. Fruits characteristics -such as size, shape, and color, as well as all types of cymes, and flowering days - contributed to the discrimination of the accessions. Indeterminate plants and red fruits showed higher ${ }^{\circ}$ Brix than semi-determined plants and orange, yellow, and purple fruits; however, the latter had a better flavor. A new type of tomato leaf that had not been previously reported among the tomato descriptors was found; the leaf was described as "with sprout".


Key words: Solanum lycopersicum L., native populations, morpho-agronomic characterization.

## INTRODUCTION

Tomato (Solanum lycopersicum L.) is considered the most important vegetable at domestic and international level (FAOSTAT, 2020). Currently, Mexico is the main exporter of tomato. The country holds a $26.5 \%$ share of the market, out of which $99.8 \%$ is exported to the USA, amounting to US $\$ 2,601,163,000$ (TRADE MAP, 2021). Mexico is considered as the center of domestication and genetic diversity of tomato (Peralta and Spooner, 2007). Recent studies confirm that tomato crops have lost genetic variability, as a result of the constant selection carried out by domestication processes, promoting autogamy (Chen and Tanksley, 2004). Additionally, other factors that have influenced autogamy include the genetic improvement of specific features, such as: higher productivity, shelf life, self-pruning, plant height, precociousness, and adaptation to different cultivation systems (Bai and Lindhout, 2007). Meanwhile, the fruit quality attributes have been left aside (Klee and Tieman, 2018). This reduced genetic base has made tomato crops very sensitive to biotic and/or abiotic stress; therefore, efforts have been made to recover and preserve wild germplasms, because they have a wide genetic variability, as a result of the extreme conditions they have endured during long periods, in their agroecology environment. Consequently, these plants constitute a genetic heritage of new genes of interest. They can also be used to recover lost genes which could take part of an introgression in modern cultivars - through conventional and/or biotechnological techniques. Likewise, they can help to develop new varieties capable of facing climate change, the new challenges posed by productive systems, and the market demand for innocuous products, which must have higher sensorial and nutritional values. Therefore, the aim of this study was to evaluate the variability of wild tomato populations, based on their morphological traits, with their immediate accession in time and space in mind, in order to develop a program for the improvement of the genetic features of agronomic and commercial interest.

## MATERIALS AND METHODS

## Biological material

Fifteen wild tomato populations samples (from different areas of Mexico) and two commercial varieties (control) were evaluated (Table 1).

## Experimental design and agronomic management

A completely randomized experimental design, with 17 treatments ( 15 wild tomato populations and 2 commercial tomato populations) and 10 repetitions (individuals), was used in the experiment. The experimental unit was one plant per bag. The plants were transplanted 35 days after the sow (dds) in a hydroponic system; there was 40 cm of separation between them and 1 m between rows. The nutrient solution proposed by Sánchez and Escalante (1989) was used. The plants were put under greenhouse conditions, with a $21-24^{\circ} \mathrm{C}$ temperature and $60-70 \%$ relative humidity.

## Evaluated variables

The characterization was based on 65 morpho-agronomic descriptors, proposed by Biodiversity International (1996): 19 vegetative types and 46 reproductive types, and 29 quantitative and 36 qualitative types.

## Statistical analysis

The quantitative data were subject to an analysis of variance and a mean comparison test (Tukey, $\mathrm{p} \leq 0.05$ ), using the SAS statistical software (version 9.4); in addition, the Pearson coefficient was estimated and a correlation matrix was developed to carry out a main component analysis, using the RStudio software (version 4.0.4). A description of the features of the qualitative data was carried out and the result was subjected to a multiple correspondence analysis, using the RStudio software (version 4.0.4).

## RESULTS AND DISCUSSION

All the wild tomato populations showed a high phenotypic variability during the vegetative and reproductive stages (Table 1). The results were: $53.33 \%$ had an indetermined habit, while $46.67 \%$ showed a semi-determined habit.

The cymes of semi-determined plants were mostly finished in flower (71.43\%) and the cymes of indetermined plants reverted to a vegetative shape ( $87.50 \%$ ) with leaves

Table 1. Qualitative traits of tomato populations.

| POP | Accessions | GH | LT | TI | SP | FS | FSM | FF | Fruit type | Fruit colour |
| :---: | :--- | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | LBCh 76 | S | S | VL | S | S | I | VF | Ball | Yellow |
| 2 | LBCh 231 | S | S | EF | S | S | I | I | Ball | Red |
| 3 | LBCh 301 | S | S | EF | I | R | S | VF | Ball | Orange |
| 4 | LBCh 188 | I | S | VS | S | R | V | S | Cherry | Red |
| 5 | LBCh 86 | S | S | EF | S | H | V | F | Grape | Red |
| 6 | LBCh 82 | S | S | VS | S | R | V | VS | Ball | Red |
| 7 | LBCh 75 | I | S | VS | E | F | S | VS | kidney | Red |
| 8 | LBCh 71 | I | W | VS | S | R | V | I | Cherry | Yellow |
| 9 | LBCh 67 | I | W | VS | S | S | V | I | Cherry | Orange |
| 10 | LBCh 61 | I | S | VL | E | F | S | VS | kidney | Red |
| 11 | LBCh 2da 28 | I | W | VS | S | R | V | F | Cherry | Orange |
| 12 | LBCh 2da 18 | I | S | VS | S | F | S | VS | kidney | Red |
| 13 | LBCh 2da 11 | S | P | EF | S | S | S | I | Ball | Purple |
| 14 | LBCh 2da 02 | I | S | EF | S | C | V | F | Grape | Orange |
| 15 | LBCh 2da 09 | S | P | EF | S | S | S | I | Ball | Purple |
| 16 | Rio Grande | D | S | EF | S | H | I | I | Saladette | Red |
| 17 | Floradade | D | S | EF | S | S | I | I | Ball | Red |

POP=population; $\mathrm{GH}=$ growth habit, $\mathrm{S}=$ semi-determinate, $\mathrm{I}=$ indeterminate, $\mathrm{D}=$ determinate; $\mathrm{LT}=$ leaf type, $\mathrm{S}=$ standard, $\mathrm{W}=$ with sprout, $\mathrm{P}=$ potato leaf; $\mathrm{TI}=$ terminal meristem of inflorescence, $\mathrm{EF}=$ ended in flower, $\mathrm{VL}=$ vegetative reverted to leaf, $\mathrm{VS}=$ vegetative reverted to sprout; $\mathrm{SP}=$ stigma position, $\mathrm{S}=$ same level as anthers, $\mathrm{I}=$ inserted, $\mathrm{E}=$ exserted; $\mathrm{FS}=$ fruit shape, $\mathrm{F}=$ flattened, $\mathrm{S}=$ slightly flattened, $\mathrm{R}=$ rounded, $\mathrm{H}=$ high rounded, $\mathrm{C}=$ cylindrical; $\mathrm{FSM}=$ fruit size at maturity, $\mathrm{I}=$ intermediate, $\mathrm{S}=$ small, $\mathrm{V}=$ very small; $\mathrm{FF}=$ fruit firmness, $\mathrm{VF}=$ very firm, $\mathrm{F}=$ firme, $\mathrm{I}=$ intermediate, $\mathrm{S}=$ soft, $\mathrm{VS}=$ very soft. Rio Grande y Floradade correspond to control varieties.
and sprouts, after a certain number of flowers were formed. Most populations showed flowers with stigma at the same level of the stem cone; however, some were also slightly or excessively projected. These results match the change of position from exserted to inserted stigma during domestication, which favored self-fertilization (Chen and Tanksley, 2004); however, the inserted or same-level stigma are more common in modern materials (Blanca et al., 2012).

The most frequent type of leaves was the standard ( $66.67 \%$ ), followed by the leaves "with sprouts" (20.00\%) (Figure 1) -which had not been reported among the tomato descriptors - and, to a lesser degree, potato leaves ( $13.33 \%$ ) - which are related to high anthocyanin populations.

These results differ from those obtained by Agudelo et al. (2011), who reported a greater frequency of potato leaves ( $69.56 \%$ ) than standard leaves (30.43\%). Blanca et al. (2012) pointed out that standard leaves prevail in the cultivated species.

Most of the fruits were very small ( $46.67 \%$ ) (Figure 2) and had rounded, slightly flattened, and oblong-elongated shapes (indetermined plants) or rounded and roundish-elongated shapes (semi-determined plants). There were also small fruits (40\%) with flattened shapes (indetermined plants) and slightly flattened and rounded shape (semi-determined plants). To a lesser degree, there were medium-sized fruits ( $13.33 \%$ ), with slightly flattened shapes (semi-determined plants). Therefore, slightly flattened (33.33\%) and rounded (33.33\%) shapes were most frequent than flattened ( $20 \%$ ), roundish-elongated ( $6.67 \%$ ), and oblongelongated ( $6.67 \%$ ) shapes. One of the major consequences of domestication is the increase of fruit size (Díez and Nuez, 2008).

The populations with an indetermined growth habit bore red, orange, and yellow fruits; additionally, semi-determined plants bore more purple fruits (Figure 2). The native tomato populations of Mexico have a high variability in fruit size, shape, and color (Lobato-Ortiz et al., 2012). Consequently, the color and pigment content of tomato fruits would be a very interesting area of research that would improve its nutraceutical quality and/or meet the preferences of the consumers.


Figure 1. Leaves "with sprouts" in tomato populations 8, 9, and 11.


Figure 2. Size, shape and color of fruits for seventeen populations. Numbers indicate tomato populations. The bar ( 5 cm ) indicates a reference measurement.

Additionally, populations with high fruit consistency were found. On the one hand, after 20 days, the consistency of populations 5,3 , and 14 reached a medium point and turned soft after 40 days; likewise, the consistency of population 1 changed to medium after 60 days. On the other hand, the consistency of populations 13 and 15 -which were pigmented with anthocyanin - turned soft after 20 days; nevertheless, the integrity of their epidermis remained constant up to 30-40 days of shelf life. Control and other wild varieties showed an opposite behavior. Bonilla-Barrientos et al. (2014) reported a higher frequency of hard fruits (pepper-type) than medium fruits (cherry tomatoes) and soft fruits (kidneyshaped); these results are very similar to those obtained in this study.

## Analysis of variance

There were significative differences ( $\mathrm{p} \leq 0.05$ ) in all the evaluated variables. All the wild populations showed high phenotypic variability; however, the characteristics that helped to achieve a better discrimination were fruits and cymes (Table 2). Precocious materials and flowering were detected in populations $12,8,9,14$, and 7 ( $45-48 \mathrm{dds}$ ) and in populations 6 , 2 , and 3 (49-51 dds). Other populations behaved similarly to control -such as populations $5,10,15$, and 13 ( $55-62$ dds). Regarding fruit ripening, the populations were classified as early ( 8 and 9 ); medium, before control ( $12,14,7$, and 6 ); and medium, similar to control (2 and 3 ); medium, after control ( $5,10,15,13,4$, and 11 ); and late ripening (1).

Table 2. Quantitative traits of tomato populations.

| POP | FT | FRT | HC | NFI | IL | FW | NL | PD | ED | NSF | TSS |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | 73 | 105 | 44.75 a | 11.70 b | 46.60 b | 71.76 bc | 2.70 cd | 4.70 b | 5.50 ab | 109.80 b | 6.90 cdef |
| 2 | 50 | 86 | 30.19 b | 7.00 b | 12.94 b | 105.67 b | 3.50 bc | 5.00 a | 6.10 a | 102.70 bc | 8.10 abc |
| 3 | 51 | 85 | 20.50 bcd | 17.70 b | 30.25 b | 24.12 efg | 2.00 d | 3.40 bcd | 3.50 cdef | 66.40 cde | 8.10 abc |
| 4 | 64 | 80 | 21.24 bcd | 7.10 b | 16.85 b | 1.49 g | 2.00 d | 1.30 e | 1.40 g | 40.40 efg | 9.70 a |
| 5 | 55 | 78 | 29.70 b | 78.30 a | 117.10 a | 7.92 fg | 2.00 d | 3.10 bcd | 2.00 efg | 23.60 fg | 9.50 ab |
| 6 | 49 | 80 | 15.70 cd | 7.50 b | 20.30 b | 11.26 fg | 2.00 d | 2.50 cde | 2.70 defg | 67.00 cde | 9.00 abc |
| 7 | 48 | 80 | 22.50 bcd | 12.20 b | 33.50 b | 44.31 de | 6.90 a | 3.20 bcd | 5.00 bc | 149.50 a | 8.10 abc |
| 8 | 46 | 62 | 24.00 bcd | 6.00 b | 11.70 b | 2.43 g | 2.00 d | 1.60 de | 1.60 fg | 52.90 def | 8.00 abc |
| 9 | 46 | 64 | 21.00 bcd | 5.90 b | 17.25 b | 3.56 g | 2.00 d | 1.70 de | 1.90 efg | 52.10 def | 8.00 abc |
| 10 | 60 | 87 | 44.10 a | 8.20 b | 28.30 b | 53.90 cd | 7.00 a | 3.40 bcd | 4.80 bc | 62.70 cde | 7.90 bcd |
| 11 | 64 | 85 | 12.13 d | 12.38 b | 9.75 b | 1.37 g | 2.00 d | 1.30 e | 1.30 g | 41.40 efg | 7.80 bcd |
| 12 | 45 | 81 | 17.25 bcd | 12.13 b | 30.88 b | 27.16 def | 7.40 a | 2.50 cde | 4.20 bcde | 53.40 cdef | 9.00 abc |
| 13 | 62 | 80 | 22.88 bcd | 16.13 b | 45.75 b | 36.82 def | 2.00 d | 3.80 bc | 4.20 bcde | 41.30 efg | 5.60 defg |
| 14 | 47 | 82 | 24.70 bcd | 74.60 a | 146.40 a | 8.00 fg | 2.00 d | 3.50 bcd | 2.20 efg | 19.90 g | 9.10 abc |
| 15 | 60 | 81 | 28.30 bc | 17.60 b | 48.90 b | 42.69 de | 2.00 d | 3.90 bc | 4.40 bcd | 84.70 bcd | 5.50 efg |
| 16 | 57 | 84 | 22.50 bcd | 7.70 b | 16.85 b | 76.33 bc | 2.30 d | 6.00 a | 5.10 bc | 65.70 cde | 4.70 fg |
| 17 | 60 | 90 | 22.30 bcd | 6.20 b | 8.15 b | 145.27 a | 4.30 b | 5.80 a | 6.80 a | 176.70 a | 4.20 g |
| Media | 55 | 82 | 24.93 | 18.14 | 39.04 | 39.06 | 3.18 | 3.34 | 3.69 | 71.19 | 7.60 |
| C. V. | - | - | 32.40 | 73.97 | 68.08 | 44.24 | 23.02 | 32.11 | 28.17 | 28.96 | 20.52 |
| HSD | - | - | 13.20 | 22.11 | 42.33 | 26.74 | 1.15 | 1.84 | 1.75 | 32.42 | 2.51 |

$\mathrm{POP}=$ population; $\mathrm{FT}=$ flowering time (days); FRT=fruit ripening time (days); HC=height of the first fruit cluster (cm); NFI=number of flowers per inflorescence; $\mathrm{IL}=$ inflorescence length (cm); FW=fruit weight (g); NL=number of locules per fruit; $\mathrm{PD}=$ polar diameter of fruit $(\mathrm{cm}) ; \mathrm{ED}=$ equatorial diameter of fruit $(\mathrm{cm}) ; \mathrm{NSF}=$ number of seeds per fruit; $\mathrm{TSS}=$ total soluble solids $\left({ }^{\circ} \mathrm{Brix}\right) ; \mathrm{C} . \mathrm{V} .=$ coefficient of variation; $H S D=$ Tukey's honestly significant difference test. Letter indicate significant differences between the means $(\mathrm{P} \leq 0.05)$.

The characterization of wild materials and semi-domestic plants shows a high diversity in days to the beginning of flowering (Carrillo and Chávez, 2010) and ripening time of the fruit (Chávez-Servia et al., 2011). On this regard, Mejía- Betancourt (2020) pointed out that precocious and compact modern materials are very helpful, as a result of the efficient use of greenhouse space and time, handling high sow densities, shortening the crop cycle, and obtaining a higher number of cycles per year. Additionally, they help to reduce the production costs, as a result of the reduced use of phytosanitary and nutrimental supplies.

For practical purposes, the populations were grouped in four categories, based on the height of the first bunch. The first group was formed by populations 1 and $10(>40 \mathrm{~cm})$; the second included populations 2,5 , and $15(28-31 \mathrm{~cm})$; the third included populations 14 , $8,13,7,16,17,4,9,3$, and $12(17-25 \mathrm{~cm})$; finally, the fourth included populations 6 and 11 ( $12-16 \mathrm{~cm}$ ).

These results are similar to those obtained by Bonilla-Barrientos et al. (2014) in pepper, cherry, and kidney-shaped native varieties, which reached a $1.96-45.41 \mathrm{~cm}$ height. We must highlight those materials with bunches at very low heights ( $<20 \mathrm{~cm}$ ) suffer disadvantages, because they can be impacted by pathogens in open field production systems. This is not
an undesirable quality in intensive greenhouse crop systems, where making the best of the space, reducing the crop cycle, and obtaining higher yields is fundamental.

The flower quantity was directly related to the cyme length ( $\mathrm{r}=0.95$ ) and the cyme type. Populations 5 and 14 stood out with 4.7 bunches in average, a $>115 \mathrm{~cm}$ length, and $>70$ flowers per cyme; the rest had simple, bifurcated, and trifurcated bunches, a 9-49 cm length, and 5-18 flowers per cyme, depending on the population. Other studies have found a high variability in cherry tomatoes regarding the number of flowers - from 7.4 to 177 (Boada et al., 2010)- and cyme length -from 58.5-77.6 cm in progenitors to 147.3 cm in hybrids (Yanokuchi et al., 1994).

For practical purposes, the populations were regrouped according to fruit weight. The group with highest weight included populations 1, 2, and control ( $72-145 \mathrm{~g}$ ); the medium group included populations $3,12,13,15,7$, and $10(24-54 \mathrm{~g})$; and, finally, the lower group included populations $11,4,8,9,5,14$, and $6(1-11 \mathrm{~g})$. Chávez-Servia et al. (2011) obtained similar results using wild and semi-domesticated materials, recording $5.6-128.7 \mathrm{~g}$ per fruit.

The populations were also divided in groups, according to the number of loculus per fruit. The first group included populations 7, 10, and 12 (5-10 loculus); the second group was comprised of population 2 and Floradade (3-6 loculus); the third group included population 1 and Río Grande (2 or 3 loculus); and populations $3,4,5,6,8,9,11,13,14$, and 15 made up the final group (2 loculus).

Wild species have less loculus per fruit (2-3) than modern varieties - which usually have 2.6-36 loculus, although the actual figure can range from 2 to 30 (Grandillo and Tanksley, 1996).

Polar and equatorial diameter characteristics had a positive relation $(\mathrm{r}=0.86)$, with values from 1.3 to 5.0 and 1.3 to 6.1, respectively. These results are higher than those recorded by Chávez-Servia et al. (2011) in wild and semi-domesticated populations: a 1.4 - to $3.1-\mathrm{cm}$ polar diameter and a $1.4-$ to $3.7-\mathrm{cm}$ equatorial diameter. According to the reports of Bai and Lindhout (2007), we can conclude that all populations that show high values of loculus, as well as of fruit weight and diameter, are semi-domesticated biological materials.

Regarding total soluble solids, all the wild populations obtained more ${ }^{\circ}$ Brix (5.59.7) than Rio Grande and Floradade control plants (4.7 and 4.2, respectively). Overall, indetermined plants show higher ${ }^{\circ}$ Brix ( 8.45 average) than semi-determined plants ( 7.52 average). Regarding the fruits, red tomatoes had the highest ${ }^{\circ}$ Brix ( 8.76 average), followed by yellow ( 7.45 average), and purple ( 5.55 average); however, purple tomatoes had a better flavor. Additionally, very small tomatoes had $8.73{ }^{\circ} \mathrm{Brix}$, the medium tomatoes, $7.50^{\circ} \mathrm{Brix}$, and small tomatoes, $7.37^{\circ}$ Brix. Crisanto-Juárez et al. (2010) recorded similar values for wild harvested fruits (4.5-9.3 $\left.{ }^{\circ} \mathrm{Brix}\right)$. These results prove that these materials have excellent quality features for the improvement of modern materials.

## Multiple correspondence analysis

Taking into account the 34 morpho-agronomic qualitative characteristics, the analysis showed that the first three dimensions (Dim1, Dim2, and Dim3) accounted for $42.06 \%$ of
the total phenotypic variability. However, Garzón (2011) reported that the total variation of 36 accessions of cherry tomato accounted for $76.98 \%$ of the three first dimensions. The phenotypic variability of the populations was mainly represented by the characteristics of the fruit and the cyme; Chime et al. (2017) reported similar results.

Four features with the highest contribution to the two first dimensions were selected, subsequently, the populations were classified in four groups (Figure 3, Table 3). Diml accounted for $17.17 \%$ of the variation and was represented by shape, firmness, multiparous characteristics, and style projected position; meanwhile, Dim2 accounted for $13.02 \%$ and was represented by the type of leaves, growth habit, color intensity of the hypocotyl, and fruit size and color. Group I was comprised of populations 7 (G), 10 (J), and $12(\mathrm{~L})$, which had irregular transversal shaped features (iFT), low firmness in shelf life (dFE), flattened shape (aFF), colorless epidermis (iCE), 'cat-face' appearance (pACF), slightly projected style (lPE), and multiparous bunch ( mPI ). Group II was more diverse and included populations 1 (A), $2(\mathrm{~B}), 3(\mathrm{C}), 4(\mathrm{D}), 5(\mathrm{E}), 6(\mathrm{~F}), 14(\mathrm{~N})$, and the Rio Grande $(\mathrm{P})$ and Floradade $(\mathrm{Q})$ control plants; they were characterized by a high fruit firmness in shelf life (fFE), semi-determined growth ( sHC ), and medium-sized plants (iTP). Group III included populations $13(\mathrm{M})$ and $15(\mathrm{O})$, which had the usual features for greenishpurple unripe fruits (vmCF), purple ripe fruits (mCFM), potato leaves (ppTH), and


Figure 3. Qualitative traits associated with 15 wild tomato populations. A) vectors and eigenvalues; B) biplot with Diml and Dim2.

Table 3. Qualitative characteristics of 15 wild tomato populations. A) vectors and values provided by the authors; B) biplot Diml and Dim2.

| No. | Characteristics | Code | Dim1 | Dim2 | Dim3 |
| :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | Dark yellow seed | aCS | 0.6684402 | 2.0297843 | 2.0881350 |
| 2 | Purple hypocotyl | mCH | 1.0915815 | 3.8292992 | 3.9739250 |
| 3 | Potato leaf | ppTH | 1.0915815 | 3.8292992 | 3.9739250 |
| 4 | Purple color of ripe fruit | mCFM | 1.0915815 | 3.8292992 | 3.9739250 |
| 5 | Green-purple color of immature fruit | vmCF | 1.0915815 | 3.8292992 | 3.9739250 |
| 6 | Intermediate facility to separate the fruit | iSP | 1.2132500 | 2.2585527 | 1.0104040 |
| 7 | Leaflets with wavy margin | oMF | 0.7440139 | 1.6607922 | 0.3125941 |
| 8 | Hypocotyl with high intensity staining | alH | 0.0089196 | 3.4410878 | 1.7452220 |
| 9 | lnflorescence ending in flower | fPV | 1.1645026 | 2.4048689 | 0.8925949 |
| 10 | Semi-determinate growth habit | sHC | 1.2628357 | 1.2238481 | 0.0333420 |
| 11 | Intermediate height plant | iTP | 1.2944758 | 1.4165056 | 0.1365200 |
| 12 | High firmness fruit on shelf (10 ds) | fFE | 1.3976733 | 0.6467704 | 0.9987624 |
| 13 | Fruit with slightly cleft base | IFB | 0.9743000 | 0.9013400 | 0.9734923 |
| 14 | Small size fruit | pTF | 0.7561467 | 2.6603956 | 0.7735197 |
| 15 | Intermediate foliage density | iDF | 0.3894345 | 1.8728563 | 0.0277012 |
| 16 | Gray seed | gCS | 1.1993170 | 0.7731033 | 0.1997073 |
| 17 | Multiparous inflorescence | mPI | 3.0136009 | 0.2436924 | 0.4061571 |
| 18 | Low firmness fruit on shelf (10ds) | dFE | 3.8586452 | 0.0072673 | 0.3857630 |
| 19 | Fruits of intermediate firmness at harvest | iFC | 5.1271148 | 0.0381446 | 0.2380655 |
| 20 | Fruit with irregular cross section | iFT | 5.8700385 | 0.2078257 | 0.1033894 |
| 21 | Flattened shaped fruit | aFF | 5.8700385 | 0.2078257 | 0.1033894 |
| 22 | Fruit with irregular apex scar | iCA | 5.8700385 | 0.2078257 | 0.1033894 |
| 23 | Fruit with indented apex | iFA | 5.8708385 | 0.2078257 | 0.1033894 |
| 24 | Slightly exserted style | IPE | 4.5409601 | 0.3207046 | 0.0731773 |
| 25 | Fruit with cat-face appearance | pACF | 4.5409601 | 0.3207046 | 0.0731773 |
| 26 | Large height plant | aTP | 1.5123073 | 1.6146075 | 0.5467456 |
| 27 | Fruit with colorless epidermis | iCE | 2.0324658 | 0.3782076 | 0.8280825 |
| 28 | Inflorescence ending in vegetative and/or flower | aPV | 0.9250148 | 1.9872506 | 0.6346747 |
| 29 | Indeterminate growth habit | iHC | 1.5123073 | 1.6146075 | 0.5467456 |
| 30 | Dark brown seed | oCS | 0.0386171 | 2.1683382 | 1.3910380 |
| 31 | Pericarp with intermediate intensity | ilP | 1.3232791 | 0.5456709 | 2.8052140 |
| 32 | High foliage density | aDF | 0.4023615 | 2.0551404 | 0.0500720 |
| 33 | Very small fruit size | mTF | 0.1701719 | 2.7182851 | 0.0849405 |
| 34 | Hypocotyl with low intensity staining | blH | 0.1710982 | 3.3076166 | 0.4143379 |
| 35 | Small size seed | pTS | 0.2033297 | 4.9517903 | 0.5795743 |
| 36 | Yellow color of ripe fruit | aCFM | 0.1893568 | 2.1148779 | 0.0718591 |
| 37 | Yellow pericarp | aCP | 0.4490272 | 5.2368754 | 0.4117801 |
| 38 | Leaf type with sprout | bTH | 0.3150106 | 5.5416175 | 1.2303180 |
| 39 | Intermediate firmness fruit on shelf (10 ds) | iFE | 0.2137655 | 3.7368020 | 1.5691100 |
| 40 | Green hypocotyl | vCH | 0.1599537 | 4.2070902 | 0.6804410 |
|  | Eigenvalue |  | 0.34833078 | 0.26523764 | 0.23946287 |
|  | Percent variance |  | 17.17 | 13.08 | 11.81 |
|  | Cumulative percent variance |  | 17.17 | 30.25 | 42.06 |

purple hypocotyls (mCH). Group IV was comprised of populations $8(H), 9(\mathrm{I})$, and $11(\mathrm{~K})$, which had yellow pericarps (aCP), medium firmness in shelf life (iFE), green hypocotyls (vCH), and leaves "with sprouts" (bTH).

## Main component analysis

Based on 28 morpho-agronomic quantitative characteristics, the analysis showed that the first three components (CP1, CP2, and CP3) accounted for $67.41 \%$ of the observed phenotypic variation. CP1 and CP2 contributed $55.48 \%$ and they were mainly related to fruit and cyme features; these results are similar to the findings of Carrillo and Chávez (2010) and Bonilla-Barrientos et al. (2014), who isolated a total variability of $68.5 \%$ and $77.03 \%$, respectively. CP1 and CP2 were taken into account to associate the populations, which were divided into three groups (Figure 4, Table 4). Group I was divided into two subgroups, gathering all the colors of the fruits. The first included populations $3(\mathrm{C})$, $13(\mathrm{M}), 15(\mathrm{O}), 2(\mathrm{~B})$, and Rio Grande ( P ), while the second subgroup was comprised of populations 1 (A) and Floradade (Q). Group II included populations 4 (D), 5 (E), 6 (F), 8 $(\mathrm{H}), 14(\mathrm{~N}), 9(\mathrm{I})$, and $11(\mathrm{~K})$. Finally, Group III was made up of populations $7(\mathrm{G}), 10(\mathrm{~J})$, and $12(\mathrm{~L})$.


Figure 4. Quantitative traits associated with 15 wild tomato populations. A) vectors and eigenvalues; B) biplot with Diml and Dim2

Table 4. Quantitative characteristics of 15 wild tomato populations. A) vectors and values provided by the authors; B) biplot CP1 and CP2.

| No. | Trait | Code | PC1 | PC2 | PC3 |
| :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | Seedling emergence | EP | -0.14198804 | 0.11521405 | -0.12579729 |
| 2 | Sympodium length | LS | 0.13320737 | -0.17528546 | 0.35529871 |
| 3 | Sympodium diameter | DS | 0.18321171 | 0.08319585 | 0.00757643 |
| 4 | Leaves per sympodium | HS | -0.07523910 | -0.10923667 | 0.25064273 |
| 5 | Leaflets per leaf | FH | 0.14068266 | 0.27455916 | -0.12154306 |
| 6 | Leaf length | LH | 0.11393586 | 0.09477567 | 0.20869656 |
| 7 | Leaf width | AH | 0.14493577 | 0.07362487 | 0.13757391 |
| 8 | Height of the first fruit cluster | AR | 0.17529247 | -0.02925165 | 0.31747032 |
| 9 | Flowering time | OF | 0.11597196 | 0.17869755 | -0.00578613 |
| 10 | Flowlers per inflorescence | FI | 0.08574209 | 0.08963171 | 0.36935312 |
| 11 | lnflorescence length | Ll | -0.04857321 | 0.05337954 | 0.42025007 |
| 12 | Number of petals | NP | 0.17782611 | -0.33828452 | 0.02959308 |
| 13 | Number of sepals | NS | 0.15265601 | -0.37798700 | -0.02001142 |
| 14 | Corolla diameter | DCO | 0.21288692 | 0.12456773 | 0.20616778 |
| 15 | Calyx diameter | DCA | 0.18580503 | 0.12907483 | 0.23916102 |
| 16 | Stamen length | LE | 0.18568266 | 0.23435240 | 0.06262061 |
| 17 | Number of stamens | NE | 0.16737826 | -0.35399539 | -0.06575172 |
| 18 | Total length of the pedicel | LP | 0.05029053 | -0.28628617 | 0.15991232 |
| 19 | Abscission zone length | LA | 0.22620119 | 0.13930400 | 0.08418366 |
| 20 | Ripening time | DM | 0.20394520 | 0.08432035 | 0.09869649 |
| 21 | Pedicel scar width | AC | 0.27931441 | -0.00396691 | -0.12167903 |
| 22 | Fruit weight | PF | 0.26249150 | 0.07044068 | -0.13380377 |
| 23 | Polar diameter of fruit | DP | 0.25179816 | 0.12224396 | 0.01619772 |
| 24 | Equatorial diameter of fruit | DE | 0.27961214 | -0.03989131 | -0.08083599 |
| 25 | Number of locules per fruit | LF | 0.11858295 | -0.39097791 | -0.03009091 |
| 26 | Pericarp thickness | GP | 0.26439998 | 0.10644528 | -0.02280295 |
| 27 | Columella thickness | GC | 0.25827506 | -0.12926713 | -0.12077907 |
| 28 | Number of seeds per fruit | SF | 0.21938254 | -0.05610109 | -0.17912647 |
| 29 | Total soluble solids | GB | -0.18806195 | -0.14116580 | 0.25398408 |
|  | Eigenvalue |  | 11.69699 | 4.39148 | 3.45959 |
|  | Standard deviation |  | 3.4204 | 2.0956 | 1.8800 |
|  | Percent variance |  | 40.34 | 15.14 | 11.93 |
|  | Cumulative percent variance |  | 40.34 | 55.48 | 67.41 |

## CONCLUSIONS

Wild populations showed a high phenotypic variability in the vegetative and reproductive stages; the fruit and cyme characteristics made the most important contribution to their discrimination. We discovered a type of leaf that had never been reported among tomato descriptors and called it "with sprouts". Materials with high ${ }^{\circ}$ Brix values, high firmness in shelf life, and intense red and purple colors were detected. These elements are related to
bioactive compounds with high antioxidant capacity with great potential for the genetic improvement of modern varieties.

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## REFERENGES

Agudelo, A. A. G., Ceballos, A. N., \& Orozco, F. J. (2011). Caracterización morfológica del tomate tipo cereza (Solanum lycopersicum Linnaeus). Agronomía, 19(2), 44-53.
Bai, Y., \& Lindhout, P. (2007). Domestication and breeding of tomatoes: what have we gained and what can we gain in the future? Annals of Botany, 100(1), 1085-1094. https://doi.org/10.1093/aob/mcm 150
Bioversity International. (1996). Descriptores para el tomate (Lycopersicon spp.). Recuperado el 22 de marzo de 2022, de https://www.bioversityinternational.org/fileadmin/_migrated/uploads/tx_news/Descriptores_ para_el_tomate__Lycopersicon_spp.__489.pdf
Blanca, J., Cañizares, J., Cordero, L., Pascual, L., Díez, M. J., \& Nuez, F. (2012). Variation revealed by SNP genotyping and morphology provides insight into the origin of the tomato. PLoS ONE, 7(10), e48198. https://doi.org/ $10.1371 /$ journal.pone. 0048198
Boada, H. M. Y., Mejía, R. J. L., Ceballos, A. N., \& Orozco, F. J. (2010). Evaluación agronómica de treinta introducciones de tomate silvestre tipo cereza (Solanum lycopersicum L.). Agronomía, 18(2), 59-67.
Bonilla-Barrientos, O., Lobato-Ortiz, R., García-Zavala, J. J., Cruz-Izquierdo, S., Reyes-López, D., Hernández-Leal, E., \& Hernández-Bautista, A. (2014). Diversidad agronómica y morfológica de tomates arriñonados y tipo pimiento de uso local en Puebla y Oaxaca, México. Revista Fitotecnia Mexicana, 37(2), 129-139. https://doi.org/10.35196/rfm.2014.2.129
Carrillo, R. J. G., \& Chávez, S. J. L. (2010). Caracterización agromorfológica de muestras de tomate de Oaxaca. Revista Fitotecnia Mexicana, 33(4), 1-6.
Chávez-Servia, J. L., Carrillo-Rodríguez, J. C., Vera-Guzmán, A. M., Rodríguez-Guzmán, E., \& LobatoOrtíz, R. (2011). Utilización actual y potencial del jitomate silvestre mexicano. SINAREFI: Oaxaca, México. 72 p.
Chen, K.-Y., \& Tanksley, S. (2004). High-resolution mapping and functional analysis of se2.1: A major stigma exsertion Quantitative Trait Locus associated with the evolution from allogamy to autogamy in the genus Lycopersicon. Genetics, 168, 1563-1573. http://dx.doi.org/10.1534/genetics.103.022558
Chime, A. O., Aiwansoba, R. O., Osawaru, M. E., \& Ogwu, M. C. (2017). Morphological evaluation of tomato (Solanum lycopersicum Linn.) cultivars. Makara Journal of Science, 21(2), 97-106. https://doi.org/10.7454/ mss.v21i2.7421
Crisanto-Juárez, A. U., Vera-Guzmán, A. M., Chávez-Servia, J. L., \& Carrillo-Rodríguez, J. (2010). Calidad de frutos de tomates silvestres (Lycopersicon esculentum var. cerasiforme Dunal) de Oaxaca, México. Revista Fitotecnia Mexicana, 33(4), 7-13. https://doi.org/10.35196/rfm.2010.Especial_4.7
Díez, M. J., \& Nuez, F. (2008). Tomato. In: Vegetables II, Prohens, J. \& Nuez, F., Eds.; Springer: Berlin, Germany. pp. 249-323. https://doi.org/10.1007/978-0-387-74110-9
FAOSTAT. (2020). Datos sobre alimentación y agricultura. Organización de las Naciones Unidas para la Alimentación y la Agricultura. Recuperado el 2 de marzo de 2020, de FAOSTAT: http://www.fao.org/ faostat/es/\#data/QC
Garzón, R. J. P. (2011). Caracterización y evaluación morfoagronómica de la colección de tomate tipo cherry de la Universidad Nacional de Colombia sede Palmira. Tesis de maestría, Universidad Nacional de Colombia, Palmira, Colombia. 56 p.
Grandillo, S., \& Tanksley, S. D. (1996). QTL analysis of horticultural traits differentiating the cultivated tomato from the closely related species Lycopersicon pimpinellifolium. Theoretical and Applied Genetics, 92(1), 935951. https://doi.org/10.1007/BF00224033

Klee, H. J., \& Tieman, D. M. (2018). The genetics of fruit flavour preferences. Nature Reviewes Genetics, 19, 347356. https://doi.org/10.1038/s41576-018-0002-5

Lobato-Ortíz, R., Rodríguez-Guzmán, E., Carrillo-Rodríguez, J. C., Chávez-Servia, J. L., Sánchez-Peña, P., \& Aguilar-Meléndez, A. (2012). Exploración, colecta y conservación de recursos genéticos de jitomate: avances en la Red de Jitomate. SINAREFI: Texcoco, Estado de México, México. 56 p.
Mejía-Betancourt, F. A. (2020). Manejo de esquejes enraizados para la producción de jitomate en alta densidad de población, bajo invernadero e hidroponía. Tesis de maestría. Universidad Autónoma Chapingo, Texcoco, Estado de México, México.
Peralta, E. I., \& Spooner, M. D. (2007). History, origin and early cultivation of tomato (Solanaceae). In: Genetic Improvement of Solanaceous Crops, Razdan, M. \& Mattoo, A., Eds.; Science Publishers: USA. pp. 1-24.
Rodríguez-Valdés, A., Florido-Bacallao, M., Dueñas-Hurtado, F., Muñoz-Calvo, L. J., Hanson, P., \& ÁlvarezGil, M. (2017). Caracterización morfoagronómica en líneas de tomate (Solanum lycopersicum L.) con resistencia a begomovirus. Cultivos Tropicales, 38(2), 70-79.
Sánchez, F., \& Escalante, E. (1989). Hidroponía: un sistema de producción de plantas. 3a ed.; Universidad Autónoma Chapingo: Texcoco, Estado de México, México.
Tengö, M., \& Belfrage, K. (2004). Local management practices for dealing with change and uncertainty: A cross-scale comparison of cases in Sweden and Tanzania. Ecology and Society, 9(3), 1-22. https://doi. org/10.5751/ES-00672-090304
TRADE MAP. (2021). Datos comerciales mensuales, trimestrales y anuales. Estadísticas del comercio para el desarrollo internacional de las empresas. Consultado el 12 de marzo de 2021, en TRADE MAP: https://www.trademap.org/Country_SelProductCountry.aspx?nvpm=3\|484\|\|\|\|0702\% $7 \mathrm{c} \% 7 \mathrm{c} \% 7 \mathrm{c} 4 \% 7 \mathrm{c} 1 \% 7 \mathrm{cl} \% 7 \mathrm{c} 2 \% 7 \mathrm{c} 1 \% 7 \mathrm{c} \% 7 \mathrm{c} 2 \% 7 \mathrm{c} 1 \% 7 \mathrm{c} \% 7 \mathrm{cl}$
Yanokuchi, Y., Fujino, M., Ishii, T., \& Uchiumi, T. (1994). Inheritance of the sideshootless character and the length of pedicel in cherry tomatoes. Tohoku Agricultural Research, 47, 277-278. http://www.naro.affrc. go.jp/org/tarc/to-noken/DB/DATA/047/047-277.pdf

